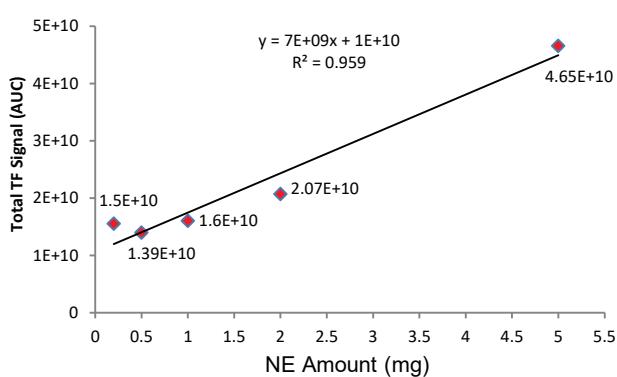
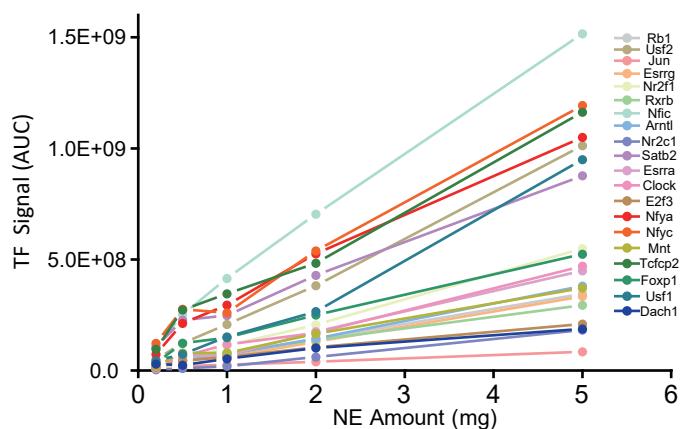
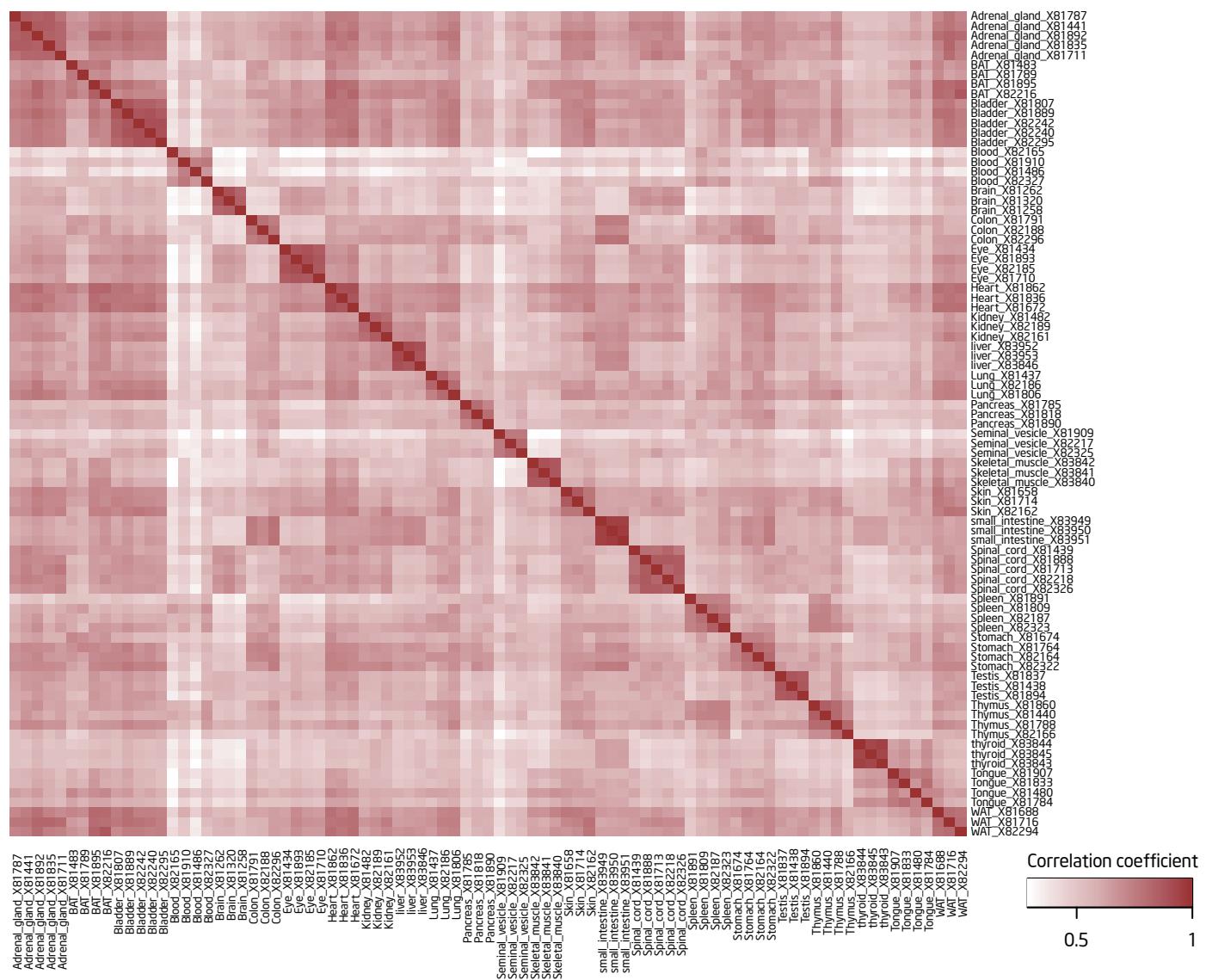
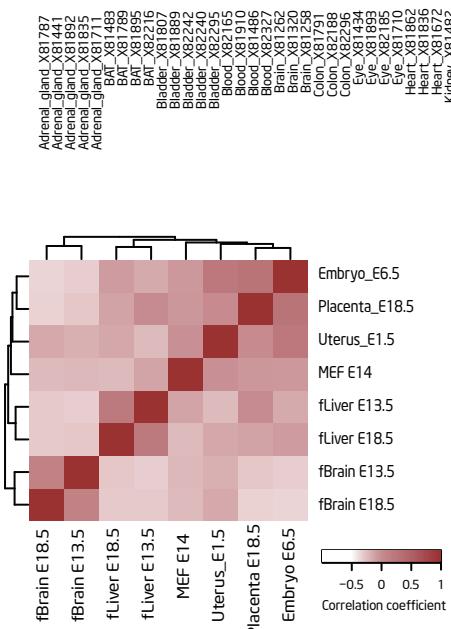
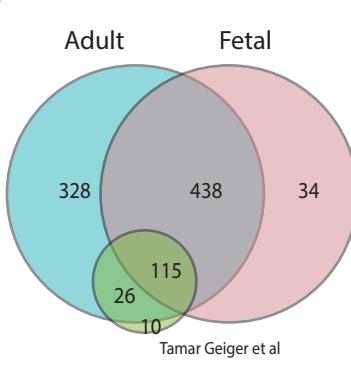
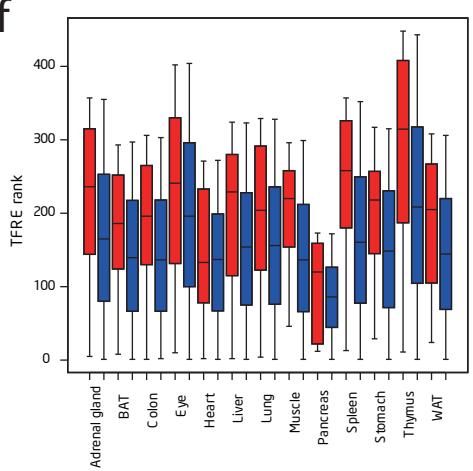
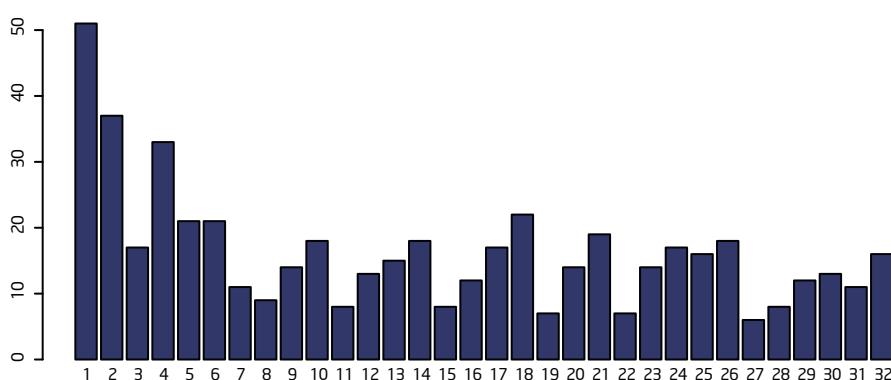


**a****b****c****d****e****f**

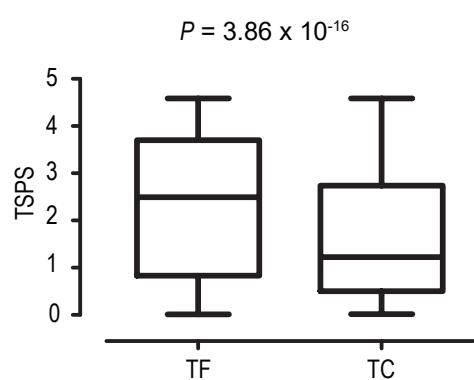
**Supplementary Figure 1. Correlation of the TF profiles between biological replicates and comparison with profiling data.**

(a) Quantitative feasibility and linearity of catTFRE strategy evaluated by dilution analysis. Different amounts of NE extracted in brain were used as shown. Total peptide AUC (area under curve) was calculated. (b) Quantitative feasibility and linearity of individual DBTFs. (c) Heat map of the TF DNA-binding abilities in the NEs (rows) from 84 samples of 24 adult mouse tissues. (d) Heat map of the TF DNA-binding abilities in the NEs (rows) from 8 fetal mouse tissues. (e) Venn diagram shows our data covers most of TFs that Geiger et al identified. (f) TFs detected in profiling data are in higher abundance part of TFRE data. Y-axis showed TF rank in TFRE data. Red boxes are overlapped TFs by profiling data and TFRE data.

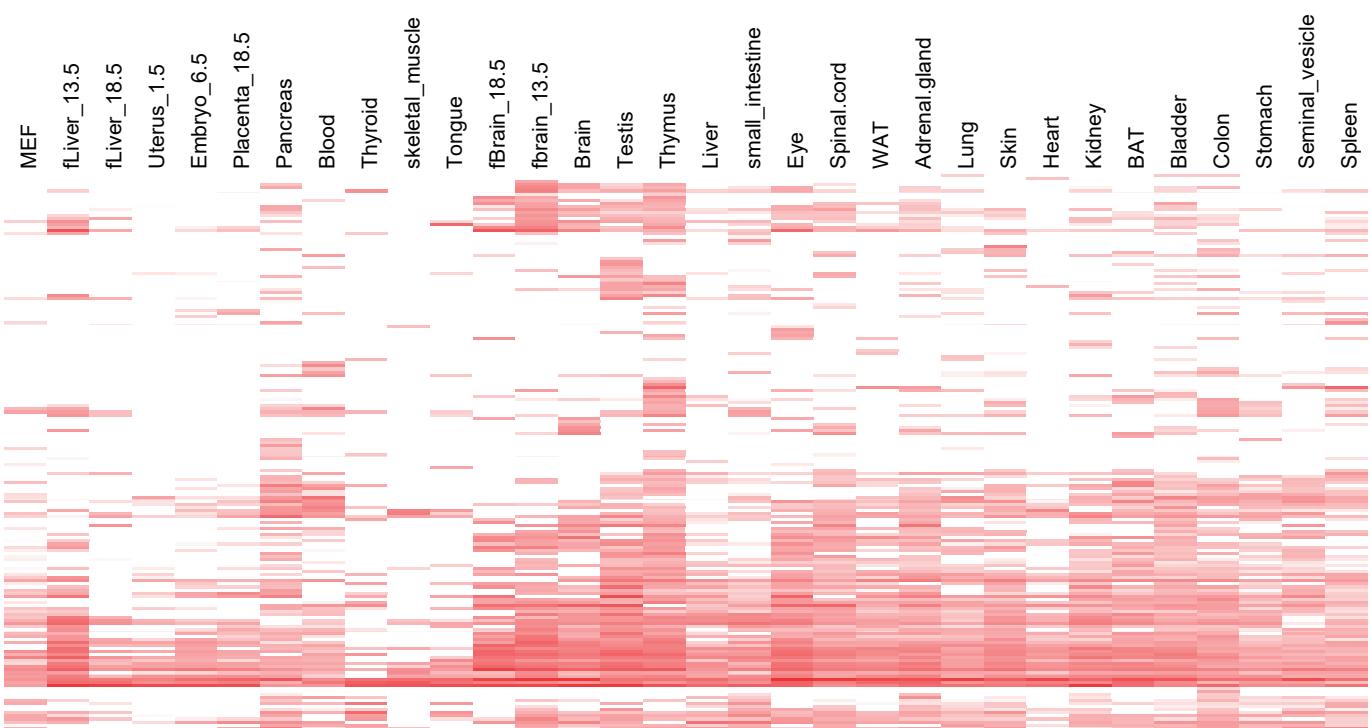
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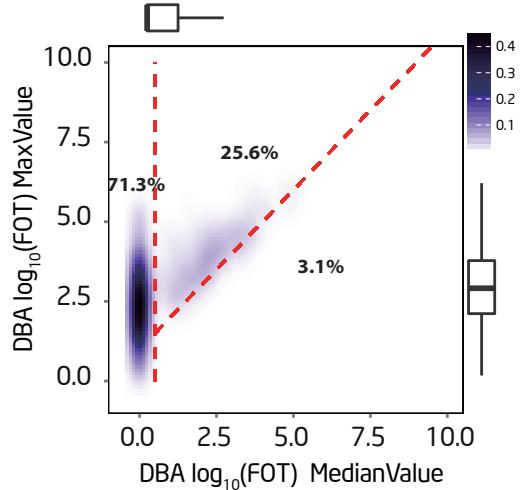
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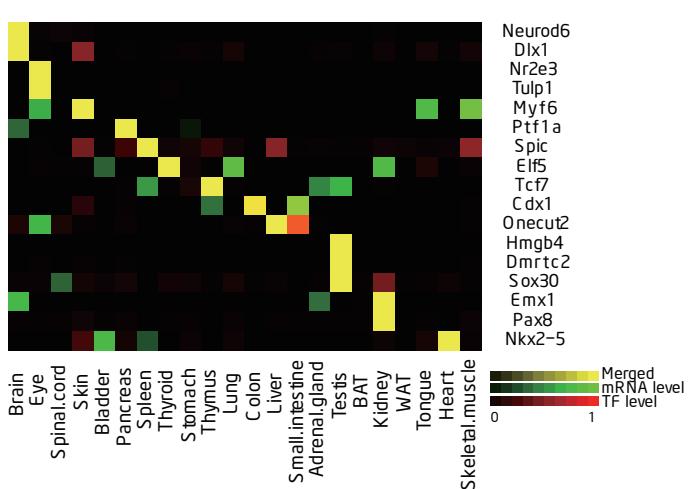
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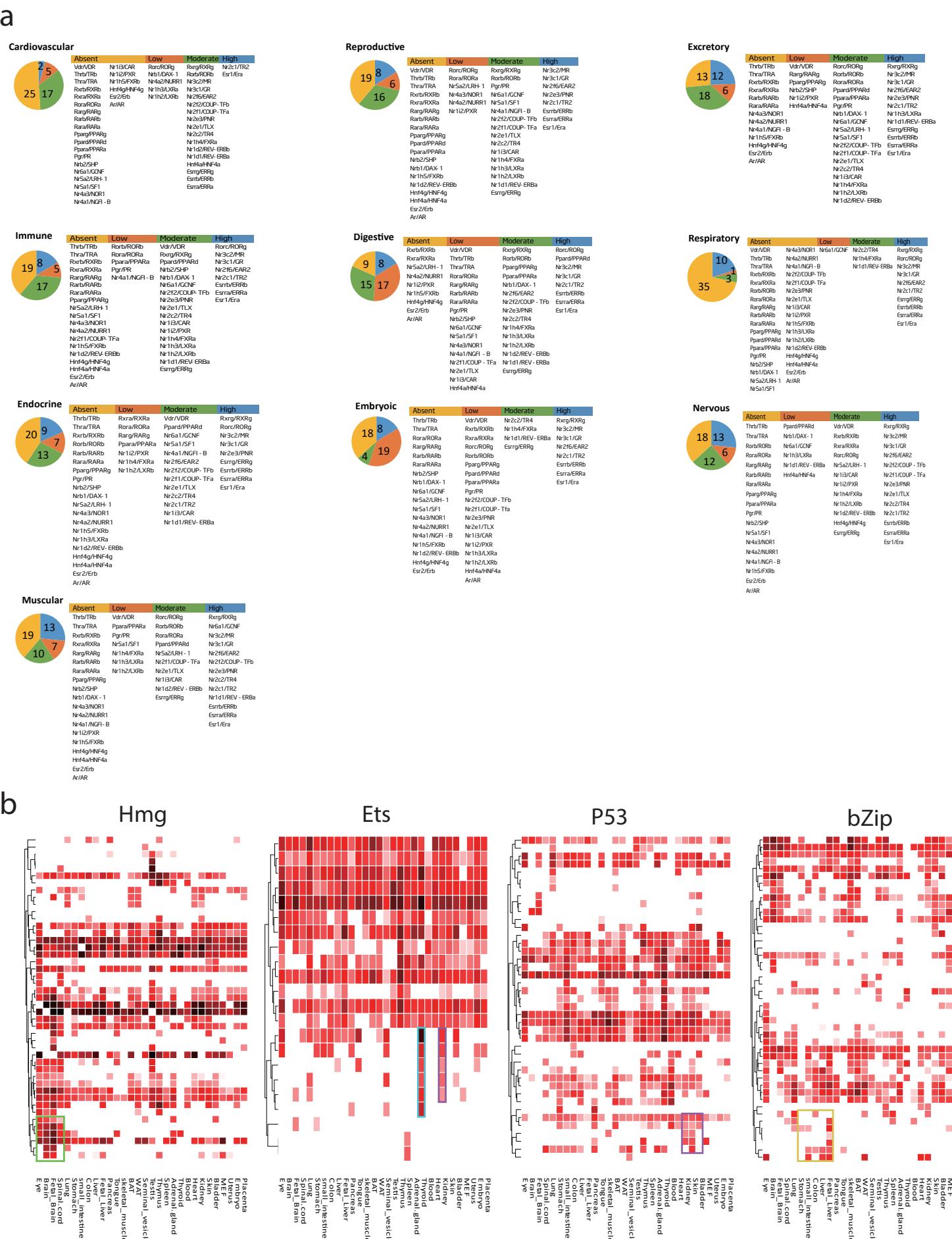


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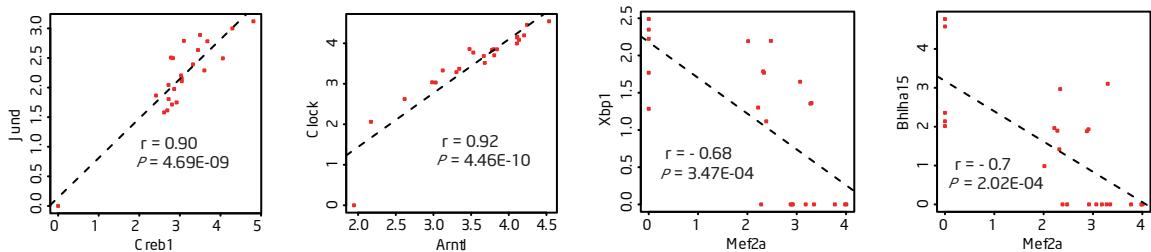
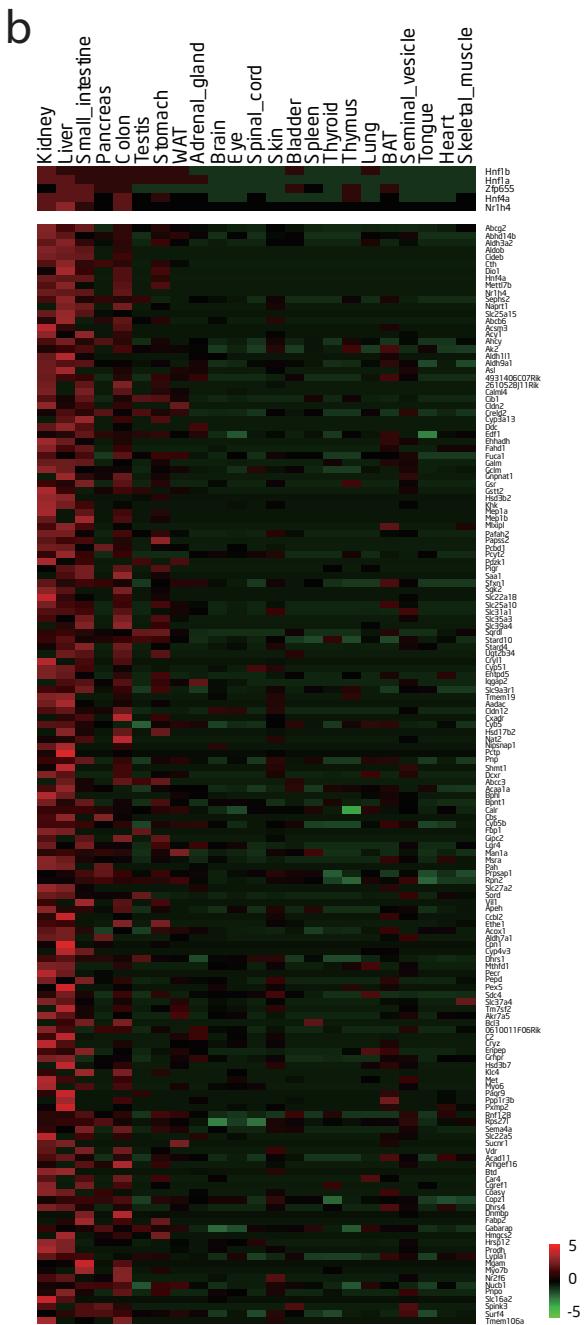
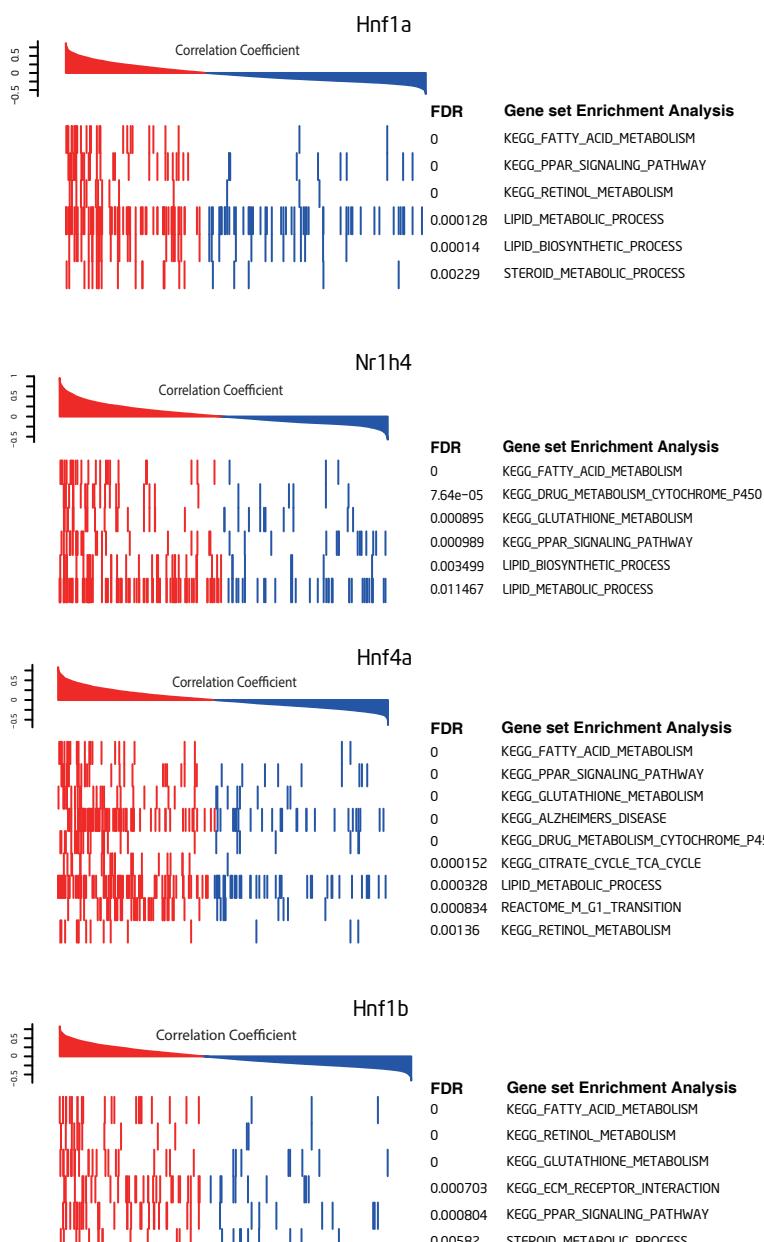
**Supplementary Figure 2. TC identifications and tissue-type restricted TF expressions in 32 mouse tissues.**

(a) Number of tissues in which the TCs are expressed. (b) Comparison of TSPS between TF and TC. (c) Heatmap for TCs in the 32 mouse tissues. (d) Distributions of ubiquitous and ttrTFs and their relative abundances in 24 adult tissues. The abundance of the TFs spans almost 7 orders of magnitude. The names of the top 3 most abundant TFs are listed. (e) Validation of 17 ttrTFs by qPCR using actin as a control in adult tissues. Red color: TF DNA binding activity; green color: mRNA; yellow color: merging of the mRNA and TF DNA binding activities.



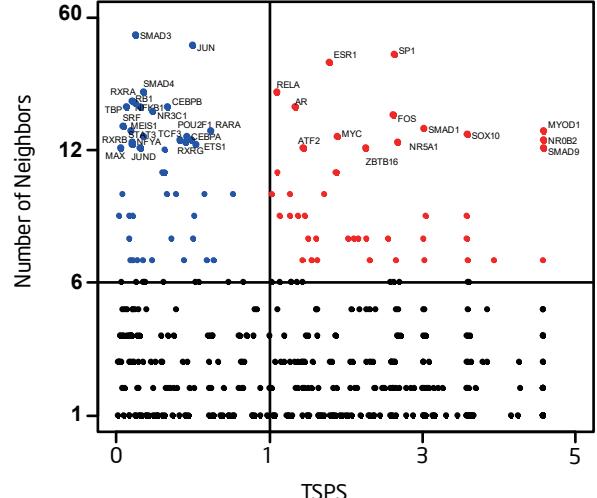
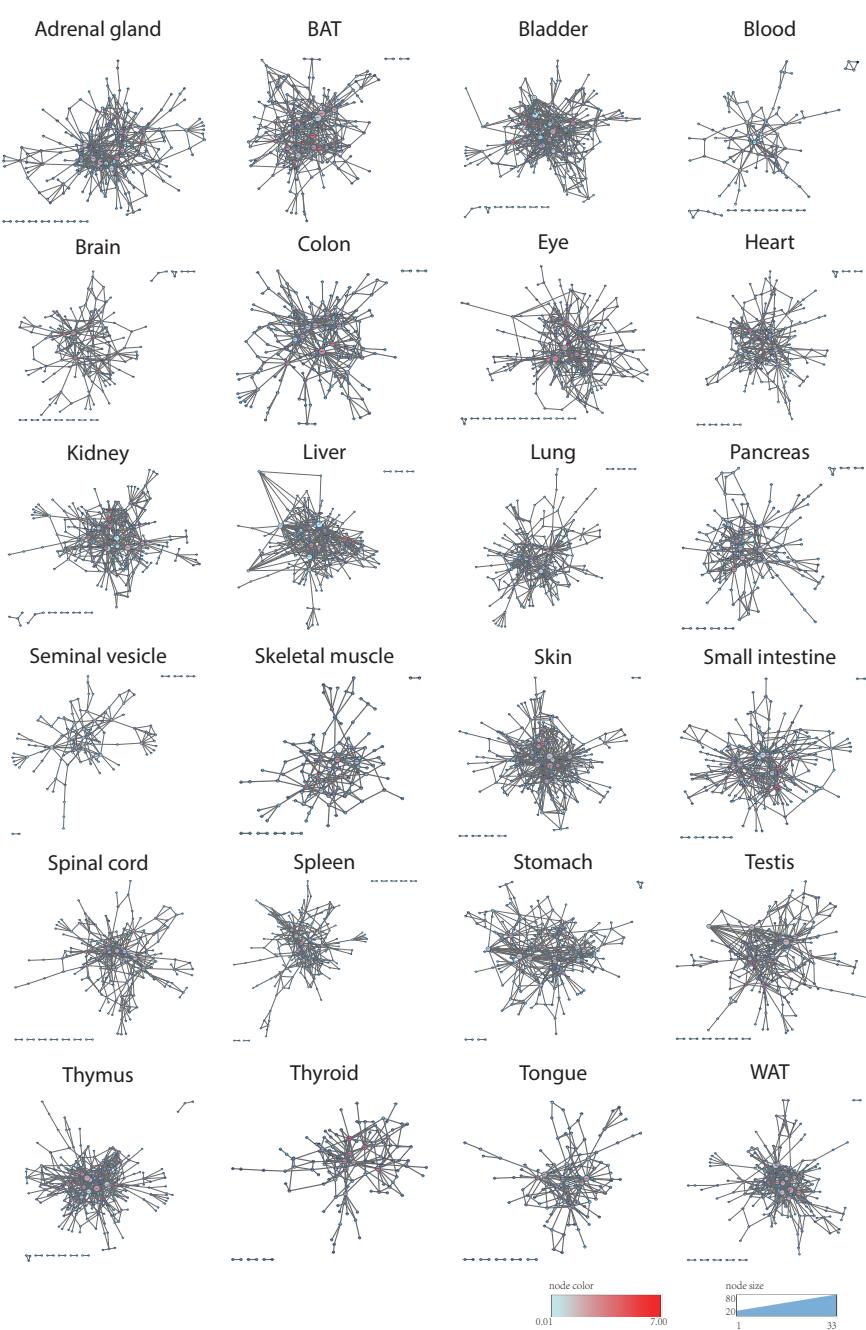
### Supplementary Figure 3. Distributions of NRs in ten physiological systems.

(a) The NR expression levels in different physiological systems are indicated by the pie charts, and their names are listed in the tables to the right. The tissue systems were categorized as described in Figure 1. (b) Unsupervised hierarchical clustering of other TF families. Blocks with different colors represent different physiological systems.

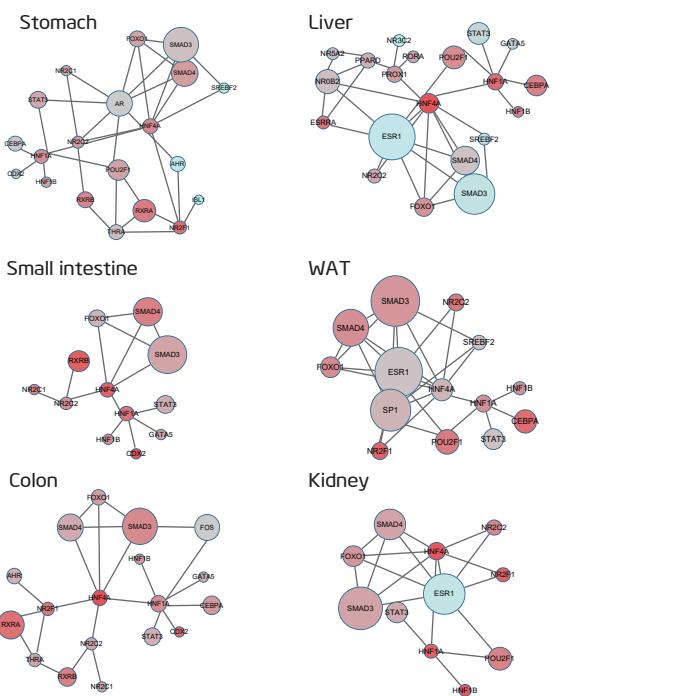


#### Supplementary Figure 4. DBTF co-expression network in 24 adult tissues.

(a) GSEA terms of other four TFs in Module #12 suggest their functions in metabolism. (b) Gene sets associated with the TFs in nine small modules. For example, Module #12 contained 5 TFs, and the mRNAs in the leading edge of the GSEA for the 5 TFs were enriched for both metabolism and immunity. (c) Correlations of individual TFs. The test for the association between paired samples was based on Pearson's product moment correlation coefficient.

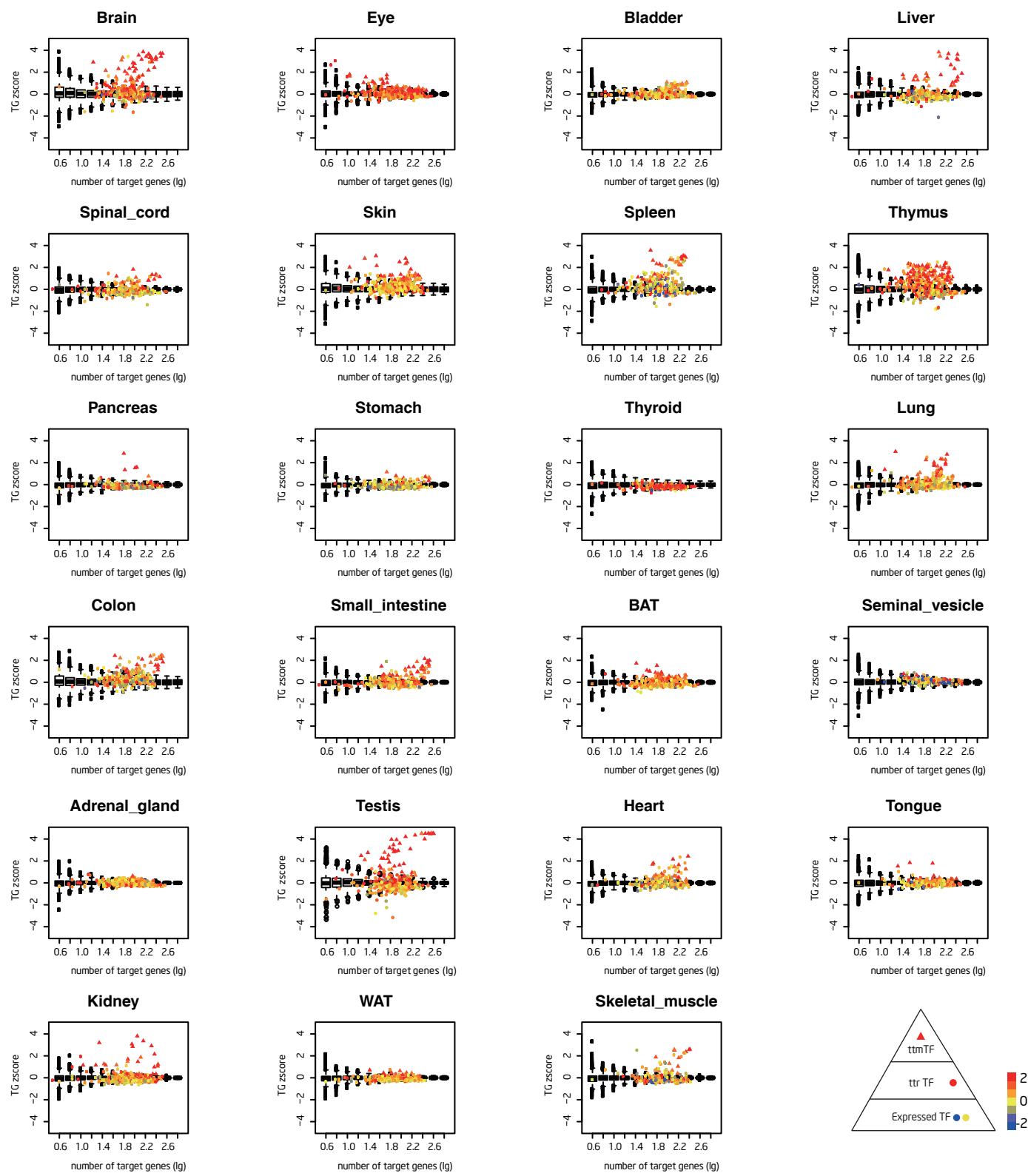
**a****b****d**

Respiratory system		Cardiovascular system	
vasculature development	1.86E-13	heart development	1.21E-16
blood vessel development	3.73E-13	cardiac muscle tissue development	1.89E-10
cell adhesion	4.20E-07	myofibril assembly	2.54E-09
angiogenesis	3.46E-07	muscle cell differentiation	1.97E-08
extracellular matrix organization	1.14E-06	actomyosin structure organization	4.74E-08
respiratory system development	2.37E-04	striated muscle tissue development	4.23E-07
morphogenesis of a branching structure	2.49E-04	muscle organ development	3.92E-07
circulatory system process	2.89E-04	cardiac cell differentiation	7.01E-06
lung development	2.89E-04		
Excretory system		Digestive system	
ion transport	2.70E-10	oxidation reduction	2.46E-12
epidermal cell differentiation	2.34E-04	steroid metabolic process	4.49E-10
transmembrane transport	3.50E-04	coagulation	5.19E-07
epidermis development	6.21E-04	regulation of body fluid levels	4.17E-07
		lipid transport	1.42E-06
		wound healing	4.38E-05
		bile acid metabolic process	1.36E-03
		complement activation	1.37E-03
Reproductive system		Endocrine system	
sexual reproduction	7.54E-72	secretion by cell	6.58E-04
gamete generation	1.50E-56	secretion	6.53E-04
reproductive process in organism	1.04E-54	antigen processing and presentation	9.76E-04
spermatogenesis	7.80E-54	synaptic vesicle transport	9.80E-04
reproductive cellular process	2.29E-27	exocytosis	6.23E-03
reproductive developmental process	6.21E-22		
meiosis	9.92E-22		
fertilization	2.37E-19		
Nervous system		Muscular system	
neuron differentiation	2.4E-03	skeletal system development	8.10E-11
transmission of nerve impulse	1.91E-02	pattern specification process	4.67E-09
spinal cord development	2.17E-02	skeletal system morphogenesis	2.39E-08
		regionalization	2.30E-08
		muscle contraction	2.81E-05
		muscle organ development	4.51E-05
		muscle system process	6.85E-05
Immune system		Embryonic tissues	
cell cycle	1.81E-41	neuron differentiation	4.80E-19
mitosis	1.15E-28	cell morphogenesis in differentiation	3.18E-09
lymphocyte activation	2.06E-16	cell part morphogenesis	3.18E-09
leukocyte activation	6.54E-16	cell projection organization	3.13E-09
positive regulation of immune system process	1.02E-13		
immune response	4.19E-10		

**c**

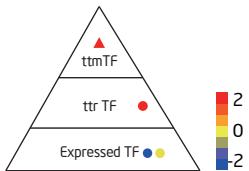
**Supplementary Figure 5. TF interaction network and functions of differentially expressed systemic TFs.**

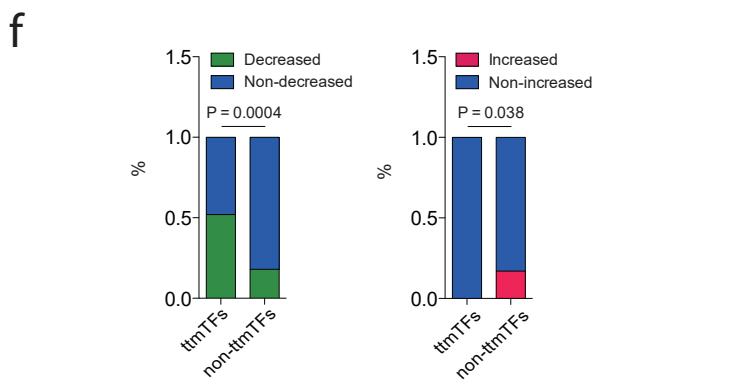
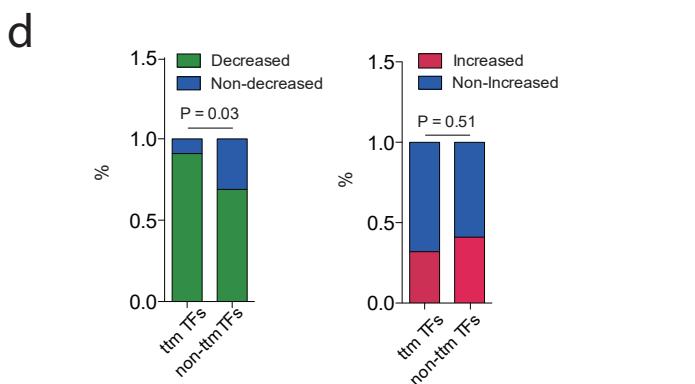
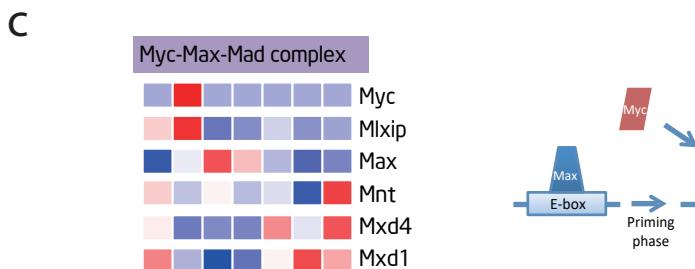
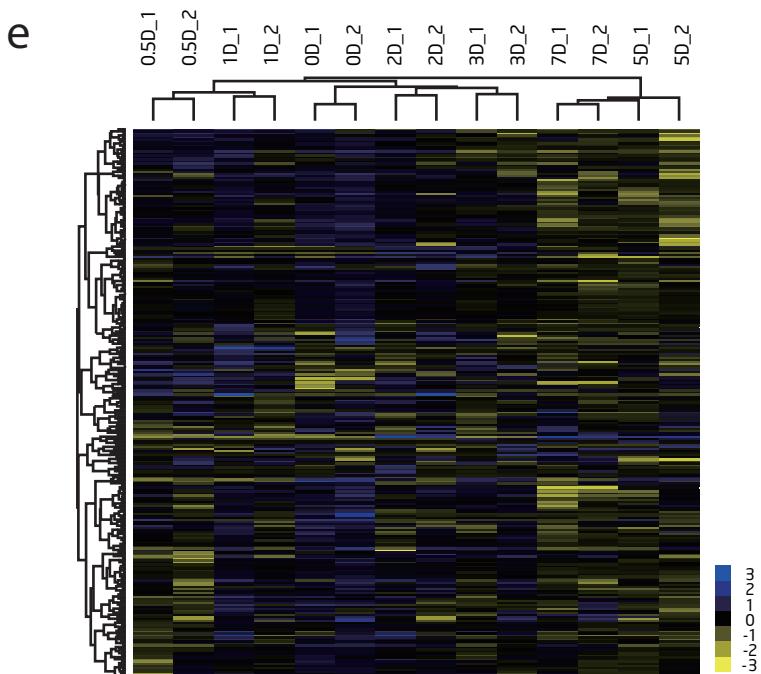
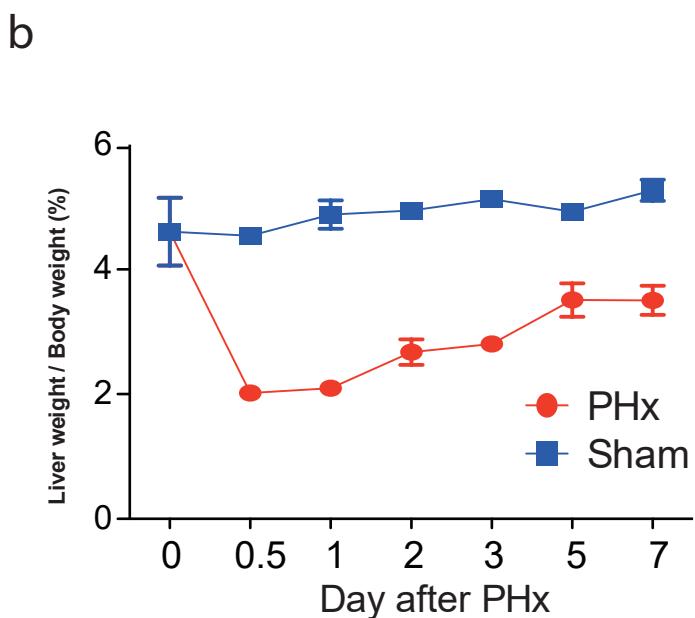
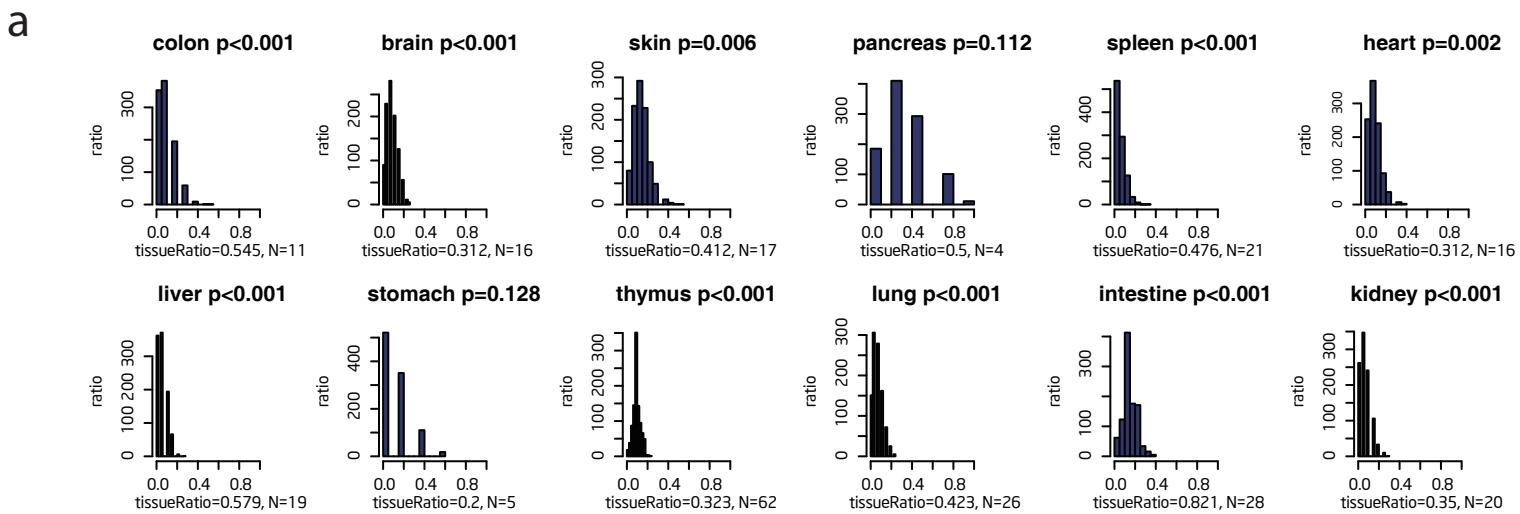
(a) Scatterplot of TSPS (x-axis) versus the number of neighbors (y-axis). Red points are defined as specifier hubs, and blue points are defined as facilitator hubs. (b) PPIs of all TFs in the 24 tissues. The colors show the DNA-binding abilities in the NEs, and the node size indicates the number of PPIs of the TF. (c) Hnf4a TF interaction network in stomach, liver, small intestine, WAT, colon and kidney. (d) The GO terms in each system were enriched using the genes that were regulated by two or more differentially expressed systemic TFs.



**Supplementary Figure 6. ttmTFs for each tissue.**

ttmTFs are TFs with high Z-scores ( $>1$ ) and TG Z-scores that are higher than those for random data.





**Supplementary Figure 7. The tsDMRs of ttmTF promoters and additional details for 2 biological repeats of the liver PHx time course data.**

(a) The ttmTF promoters in a certain tissue were enriched in that tissue's tsDMRs. Almost all tissues had an enrichment p value <0.05, except for brain ( $p=0.104$ ), kidney ( $p=0.11$ ) and heart ( $p=0.106$ ). (b) Liver/body weight ratios for 2 biological repeats of the liver PHx time course data. Data are represented as mean  $\pm$  SEM. (c) Myc/Max/Mad network in regulating liver regeneration. (d) Enrichment of down-regulated and up-regulated ttmTFs after PHx. The ttmTFs were significantly decreased compared with the non-ttm TFs after PHx. Fisher's exact test was used to test for significance. (e) Hierarchical clustering of the TF patterns during liver regeneration. Two biological repeats for each time point were co-clustered. (f) TF's target genes in profiling data were used to predict change in TFs. And enrichment of down-regulated ttmTFs was also detected in the PHx time-course data, while no enrichment for up-regulated ttmTFs. Fisher's exact test was performed to test for significance.